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EUTRALIZING MONOCLONAL ANTIBODY 2FS	ROSS-N	(54) Tile: Fab-EPITOPE COMPLEX FROM THE HIV-1 CROSS-NEUTRALIZING MONOCLONAL ANTIBODY 2F5	
		(74) Agent: STEWART, Michael, I.; Sim & McBurney. 6th floor. 330 University Avenue, Toronto, Omario M5G 1R7 (CA).	
	mig.	(72) Inventors; and (75) Inventors, Applicants (for US only): PAI, Emil, F. (DE/CA); SI (75) Inventors/Applicants (for US only): PAI, EMIL, R. (CA), KLEIN, Duncilo Divic, Toronto, Ontario MSX 2X7 (CA), KLEIN, Michel, H. (CA/CA); 16 Memo Boulevard, Willowdale, Ontario M2P 189 (CA), CHONG, Peb (CA/CA); 23 Ed- toril Streat, Richmond Hill), Ontario LA (068 (CA), PEDY- CZAK, Arthur [CA/CA]; 1399 Colmar Avenue, Pickering, Ontario L1W 1C2 (CA).	
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constructed, as set forth herein, peptide-mimetics constrained in the same β -turnexpected to increase the immunogenicity of the epitope sequence. like configuration as seen in the crystal structure of the complex, which would be The elucidation of these three-dimensional structures enables there to be

5 determined for the crystal structure, as set forth in Table 2 below of such form of the fragment to be determined and such structure is shown in crystalline form of the Fab'2F5 fragment enables the three-dimensional structure crystal of the Fab' fragment of monoclonal antibody 2F5. The isolation of the Figure 1, described below. Certain characterizing parameters have been Accordingly, in one aspect of the invention, there is provided an isolated

Protein Data Bank under Accession No. 2F5A. another space group with its own unique cell dimensions. The crystalline form of dimensions a=63.6 Å; b=76.4 Å; c=93.4 Å, although the crystals may be grown in the Fab'2F5 may have the atomic coordinates deposited on April 9, 1999 with the The isolated crystal may be grown in space group P2,2,2, with cell

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in Table 3 below. structure as shown in Figure 4, described below and atomic coordinates as shown residues H98 to H120, as seen in Table 1 below, which has a three-dimensional possess a third hypervariable (V3) loop of the heavy chain comprising amino acid Fab'2F5 molecules organized in the isolated crystal provided herein

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The crystal structure of the Feb' fragment of Meb 2P5, a potent neutralizer of both laboratory strains and primary clinical isolates of most clades of HIV-1, both uncomplexed and complexed with the largely conserved peptide sequence ELDKWAS of the viral envelope most clades of HIV-1, both uncomplexed and complexed with the largely conserved peptide sequence ELDKWAS of the viral envelope protein gp41, has been clucidated and the characteristics of peptide-protein interactions determined. Having regard to such determination, the peptide-mimetrics are constrained in the three-dimensional structure to provide an increased immunogenicity to the epitope sequence.

(57) Abstract

25 complex enables the three-dimensional structure of such form of the complex to is shown in Figure 3, described below. Certain characterizing parameters have be determined and the detail of the binding site of the peptide to the Fab' fragment No: 1) or a functional analog thereof. The solution of the crystal form of the provided an isolated crystal of the Fab' fragment of monoclonal antibody 2F5 been determined for the crystal structure of the complex, as set forth in Table 2 complexed with a peptide having the amino acid sequence ELDKWAS (SEQ ID In accordance with a further aspect of the present invention, there is

30 may be grown in another space group with its own unique cell dimensions. The cell dimensions a=58.0 Å; b=65.0 Å; c=175.6 Å, although the crystal complex The isolated crystal complex may be grown in space group P2,2,2, with

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crystalline form of the complexed form of the Fab'2F5 may have the atomic coordinates deposited with the Protein Data Bank under Accession No. 2F5B on

in which lysine is replaced by arginine and/or one in which tryptophan is replaced functional analog is ELDRWAS (SEQ ID No: 2). tyrosine, phenylalanine or uncharged histadine. One example of such The functional analog of the amino acid sequence ELDKWAS may be one

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be the same, irrespective of the kind of crystal which is analyzed conformation of the peptide epitope when it is bound to the antibody, which will bound to the peptide ELDKWAS (SEQ ID No: 1), provides details of the actual The elucidation of the crystal structure of the Fab2F5 fragment when

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HIV vaccine. epitope then provides the basis for the provision of peptide analogs, peptide. mimetics and other antigens which are potentially useful as components of an anti-The information which is provided concerning the conformation of peptide

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synthetic peptide which binds to monoclonal antibody 2F5 and which is peptide ELDKWAS (SEQ ID No: 1) shown in Figure 3. constrained to provide a three-dimensional structure corresponding to that for the Accordingly, in another aspect of the present invention, there is provided a

20 configuration of the three-dimensional structures with the tryptophan and lysine functional analog thereof and may be constrained in the slightly distorted β-turn residue chains stacked and parallel, as seen in Figure 3 and as discussed in more This synthetic peptide may contain the amino acid sequence DKW or a

30 in Figure 3, which also binds to the Mab. For example, arginine (R) may be used substituted by an amino acid which retains the peptide-protein interaction shown indicates that at least one of the tryptophan and lysine sidechains may be (H) may be used in place of tryptophan (W). Peptides wherein one or more of such in place of lysine (K) and tyrosine (Y), phenylalanine (F) and uncharged histadine amino acid substitution is effected are peptides which contain a "functional The analysis of the three-dimensioned conformation of the epitope

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analog" of the amino acid sequence DKW, as the term is understood herein, in that

the peptide still binds to the monoclonal antibody 2F5

S used to maintain the amino acid sequence DKW or analogs thereof in the conformation by any convenient means. For example, a disulphide bridge may be peptide ECDKWCS (SEQ ID No.: 3). respective orientation of two amino acid residues as shown in Figure 3. Such disulphide bridge may be provided between cysteine residues in the synthetic The synthetic peptide provided herein may be constrained in the required

5 sequence DKW or functional analogs thereof in the respective orientation of the peptide EdapDKWES (SEQ ID No.: 4) or EEDKWDapS (SEQ ID No.: 5). between diaminopropionic acid (Dap) and glutamate (E) residues in the synthetic amino acid residues as shown in Figure 3. Such lactam bond may be formed Alternatively, a lactam bond may be used to maintain the amino acid

ᄗ conjugation to carrier molecules, such as proteins, including diphtheria toxoid, may be linked to the peptide. tetanus toxoid or an outer membrane protein of Haemophilus. Such carrier protein It is well known that the immunogenicity of peptides may be enhanced by

25 20 functional analogs thereof constrained in the determined three-dimensioned thereof to form a crystalline complex; analyzing the crystalline complex to having the amino acid sequence ELDKWAS (SEQ ID No.: 1) or functional analog crystallizing a Fab' fragment of the monoclonal antibody 2F5 with a peptide of making a peptide binding to monoclonal antibody 2F5, which comprises co-Fab' fragment; and synthesizing a peptide containing at least amino acids DKW or determine the three-dimensional orientation of the bound peptide in relation to the There is also provided, in an additional aspect of the invention, a method

is one which still binds to the monoclonal antibody 2F5. Functional analogs also extend to known analogs of the ELDKWAS motif, including those of the formula The functional analog of the peptide containing at least amino acids DKW

 X_1LDKWX_2S wherein X_1 is E, A, G or Q and X_2 is A or T.

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BRIEF DESCRIPTION OF THE DRAWINGS

The file of this patent application contains drawings executed in color, namely Figures 1 to 4. Copies of this patent with color drawing(s) will be provided by the Patent and Trademark Office upon request and payment of the processory for

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Figure 1 is a colored ribbon diagram of crystalline Fab'2F5, showing the heavy chain in purple, the light chain in blue and the elongated VH3 loop (colored in gold) extending from the protein surface, as generated by MOLSCRIPT (ref.

27) and Raster 3D (ref. 28);

Figure 2 is a colored stereoplot of the ELDKWAS peptide model in density, as generated by the program O (ref. 29). The Fo-Fc map was calculated with the peptide omitted and contoured at 3σ. A minor break in the density at P7-Ser at the contour level illustrates the slight increase in flexibility at the extremes of the bound epitope;

Figure 3 is a color representation of the antigen binding site of Fab'2F5, showing protein/peptide interactions, as generated using the program SETOR (ref. 30). The residues are colored by atom type: oxygen is red, nitrogen is blue, carbon is grey and sulfur is yellow. For clarity, some hydrophobic sidechains which interact with the epitope have been omitted. All bond lengths are given in A, and

Figure 4 is a color representation of the third hypervariable loop of the heavy chain of Fab'2F5 complex comprising amino acid residues H98 to H120, as generated using the program SETOR (ref. 30). The residues are colored by atom

GENERAL DESCRIPTION OF INVENTION

The crystalline structure of the Fab' fragment of Mab 2F5 (IgG) was solved at 2.05 Å resolution by molecular replacement and adopts the standard immunoglobulin fold. A salient feature of the structure is the elongated (22 amino acids) hypervariable loop 3 of the heavy chain (V-H3, ref. 9), which comprises amino acid residues H98 to 120 and extends away from the protein surface, as can be seen from the ribbon diagram of Figure 1. The V-H3 loop is shown in detail in Figure 4. The atomic coordinates of the V-H3 loop are given in Table 3.

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In the structure of the Fab'2F5 complex with bound epitope, refined at 2.0 Å, this loop is well-defined by clear electron density. In the uncomplexed form, while the V-H3 region is less clear, loops at the C-terminal regions of the heavy chain constant domain, including the C-termini of both chains, were better resolved. Conformations from the better-defined electron density were used as templates for building the other model. The refined models comprise residues L1 to L214 of the light chain and residues H1 to H146 and H151 to H235 of the heavy chain plus ordered water molecules. The amino acid sequences of the light chain (SEQ ID No.: 2) and heavy chain (SEQ ID No.: 3) of Fab'2F5 are shown in Table 1 below. For the structure of the complex, P1 to P7 are the residues of the peptide. The H147 to H150 loop of the constant domain of the heavy chain was not visible in either structure. (Residues are labelled herein H1 to H235 for the heavy chain and L1 to L214 for the light chain and P1 to P7 for the peptides).

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5 25 20 elbow angle in the complexed form differs from uncomplexed Fab'2F5 (142° vs. angle) rather than any modification of the antigen binding site. Superpositioning smaller. An overlay of all Ca atoms results in an rmsd of 0.7 Å, but these shifts unit cells are different, uncomplexed Fab'2F5 having a unit cell which is 30% 146°). Both of these observations may be artifacts of crystal packing, since the gp41 coiled-coil trimer of the neutralization mechanism, perhaps by disrupting the conformation of the interactions with a portion of gp41 C-terminal to the epitope sequence, enhancing analysis do not provide any obvious explanation for the long insertion in the V-H3 only the variable regions gives an rmsd of 0.4 Å. While the results of the structural binding and increasing the specificity of the Fab. It may even form an integral part suggests it plays a role in the antibody mechanism. It may be involved in loop has been identified, its unusually hydrophobic nature for surface residues appear to be the result of a concerted domain movement (i.e. the change in elbow Along with differences in mobility of the loops mentioned above, the

In the complexed structure, the ELDKWAS peptide forms a slightly distorted, type I β turn, centered between P4-Lys and P5-Trp, (as seen in Figures 2 and 3), with a 3.1 Å hydrogen bond from the amide nitrogen of P6-Ala to the carbonyl oxygen of P3-Asp. The arrangement is atypical in that neither position

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and tryptophan. The dihedral angles for P5-Trp fall in the "unfavoured" region of two or three in the turn is a glycine (ref. 10), but rather the bulky residues lysine a Ramachandran plot (φ=-101.7°, ψ=8.7°).

2 arrangement of the adjacent P5-Trp and P4-Lys sidechains, with hydrophobic and H58-Asp. While the principal hydrophobic contacts of P5-Trp are the P4-Lys hydrophobic methylene groups are sandwiched between P5-Trp and H54-Tyr, rings of P5-Trp at a distance of about 3.8 A. The lysine sidechain, whose interactions between the fully-extended alkyl chain of the P4-Lys and the aromatic methylene groups, other hydrophobic residues within 4 Å of the aromatic ring ends with a sharp turn at the final amino group, forming contacts with H56-Asp backbone amide of P5-Trp as well as to L96-His-Ne and H100-Arg-NH1, all of H113-Arg. A key component to the stability of the peptide configuration is the system include H103-Pro and H32-Phe and the methylene groups of the sidechain orientation of the P3-Asp sidechain, which forms strong hydrogen bonds to the Lys-Trp (DKW) trio are the essential component of the protein/peptide 2.7 and 2.8 Å respectively. From this analysis, it can be concluded that the Aspforms strong hydrogen bonds to backbone carbonyls of H33-Gly and H101-Arg at about 2.8 Å long. A water molecule associated with P5-Trp-Ne1 at 3.0 Å also Another interesting feature of the complexed structure is the stacked

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30 glycoprotein sequence (ref. 4). For the critical portion of the epitope, DKW, estimated that the LDKW motif is 83% conserved among HIV-1 envelope modifications within the trio usually prevent neutralization (ref. 5). It was the DKW core do not have a dramatic effect on binding, whereas major where an arginine is substituted for P4-Lys (i.e. peptide ELDRWAS (SEQ ID No: 2F5 also selected sequences with a DRW core (ref. 4). The structure of a complex clade, conservation is 92% (91/99 sequences). Phage library screening with Mab 1997 to 1998 Los Alamos HIV Sequence Database (ref. 11) is 86%. Within the B conservation among 206 sequenced HIV-1 envelope proteins of all clades in the 2)) shows identical peptide conformation and contacts as the complex reported This conclusion is supported by mutation studies in which changes outside

here with the consensus epitope. The total buried accessible surface area upon

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while those for the complex Fab2f5 + ELDRWAS are shown in Table 5. and uncomplexed Fab' determined using a probe of radius 1.4 Å (ref. 12)). The peptide coordinates of the complex Fab'2f5 + ELDKWAS are shown in Table 4 surface between the intact complex and the sum of the surface areas of the peptide formation of the complex is 1025 Å^2 (calculated as the difference in accessible

5 an unstructured coil (ref. 19). structure, the continuation of a long (37 aa) helix. In the other, the C-terminus is (ref. 14), ELD at the C-terminus of the crystallized portion adopted an α -helical sequence, although two reports (refs. 14, 19) include a partial sequence. In one HIV-1 (refs. 14 to 16) or SIV (refs. 17 to 19) gp41 do not incorporate the epitope coiled coil of the gp41 ectodomain (refs. 14 to 19). Most structural analyses of been proposed (ref. 13) which was consistent with an extension of the observed and seen in detail in Figure 3 was not anticipated. A helical conformation had The conformation of the gp41 epitope found in the complex with Fab'2F5

20 15 neighboring molecule in the crystal, making it probable that crystal packing forces (GST) by a nine amino acid linker (ref. 20). In this environment, the epitope protein, where it was connected to the C-terminus of glutathione-S-transferase had high thermal parameters, denoting flexibility. herein. In the GST-fusion structure, the epitope peptide interacted with a formed part of a series of tight turns but not the β -turn seen in the results described led to the observed conformation. The gp41 peptide portion of the structure also A conformation of the full epitope was determined as part of a fusion

25 30 elucidated herein explains the stronger immune response observed when the adopts very little or no stable secondary structure. The crystal structure of Fab 2F5 live virus, underlining the importance of the correct epitope conformation. antibodies (ref. 22) where a \beta-turn conformation might be induced, in contrast to epitope was introduced into loops of hemagglutinin (refs. 2, 21) or recombinant excellent humoral response of 2F5-like binding specificity but failed to neutralize hepatitis B virus surface antigen (ref. 8), where epitope grafting resulted in an Preliminary NMR studies have suggested that the ELDKWAS sequence

transiently, after assembly of the mature gp41/gp120 trimers on the virus The conformation of the gp41 epitope set forth herein may be adopted

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5 7 consequent absence of Mab 2F5 in the antisera of most infected patients. As well life span of the antigen would be consistent with its low immunogenicity and the fusogenic form has been proposed by several investigators (refs. 14, 23). A short determinations made herein, as well as an intermediate "unsprung" and nongp41, including the stable fusogenic form observed in the structural envelope, or possibly during the fusion process. A range of conformations for either because of low antibody concentration or the short lifetime of the antigenic passive immunization with Mab 2F5 in chimpanzees failed to neutralize HIV-1, conformation. As the only identified cross-neutralizing antibody against gp41 resulting in delayed infection and lower viral loads, but not protection (ref. 6). are b12 and 2G12 which define epitopes at the CD4 binding site and V3/V4 loops broadly neutralizing monoclonal antibodies identified to date and the only one Mab 2F5 is an important focus in HIV-1 vaccine research. It is one of only three This result was presumably due to insufficient opportunity for antibody binding, of gp120 respectively (ref. 6), but in these cases the epitopes are discontinuous with a short, continuous epitope. The other two known cross-neutralizing Mab's

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of the gp41 sequence found in the structure of Fab'2F5 could overcome the low immunogenicity of the epitope, making such a compound a useful component of a future HIV-1 vaccine. Development of a peptide-mimetic constrained to adopt the conformation

and involve both peptide and carbohydrate interactions (refs. 5, 6).

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EXAMPLES

equivalents are contemplated as circumstances may suggest or render expedient. not intended to limit the scope of the invention. Changes in form and substitution of Examples. These Examples are described solely for purposes of illustration and are complete understanding can be obtained by reference to the following specific descriptive sense and not for purposes of limitations Although specific terms have been employed herein, such terms are intended in a The above disclosure generally describes the present invention. A more

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y biochemistry, crystallography and immunology used but not explicitly described in Methods of molecular genetics, peptide-mimetics chemistry, protein

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are well within the ability of those skilled in the art. this disclosure and these Examples are amply reported in the scientific literature and

Fab'2F5 and its epitope complex. This Example shows the preparation, purification and crystallization of

column 3 cm wide, 5 cm high, providing about 30 mL bed volume. The column was washed overnight with 2 L of 20 mM Tris pH 8.0. albumin, DE52 cellulose was swollen in 20mM Tris pH 8.0 and packed into a albumin added to the solution for stability. To separate the protein from the standard pepsin protocols. F(ab'), was then stored with 1% (w/v) clinical human Intact human IAM 2F5 IgG antibody was transformed into F(ab'), using

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5 L of 20 mM Tris pH 8.0 and the conductivity and pH of the buffer, flow through The protein was loaded onto the column by pumping on at 1 to 5 mL/min, with and protein concentration were checked to ensure the protein bound to the column approximately 7 mL fractions were collected. pH 8.0) was run through the column until the OD_{200} went down to baseline and albumen binding to the column while the F(ab')2 does not. Buffer A (20 mM Tris 55 ml protein at 1.1 mg/ml concentration were dialysed against 2×4 to 5

20 25 Tris pH 8.0 + 0.2 M NaCl, to ensure no other proteins were present. The flow SDS-PAGE gels as well as by a positive antigen-catch ELISA assay targetting the The sample was confirmed to be F(ab'), by reducing and non-reducing native and through protein was concentrated, producing 5 x 500 µL of F(ab), at 23 mg/ml k-chain followed by a negative assay targetting the Fc part of a human antibody The albumin was eluted with a salt gradient of 20 mM Tris pH 8.0, 20 mM

30 pH 8.0 were added and the solution left for a further 30 minutes. The Fab' was final concentration of 10 mM in DTT. The solution was incubated at room $400~\mu L~100~mM$ DTT in 0.1 M Tris pH 8.0 were added to the 4 mL to provide a use in crystallization setups dialyzed overnight against 20 mM Tris pH 8.0 and concentrated to 10 mg/mL for temperature for an hour, 30 μL of a 500 mM iodoacetamide solution in 0.1 M Thi 200 µl of Fab' at 23 mg/mL were diluted to 4 mL with 0.1 M Tris pH 8.0.

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Crystals of uncomplexed Fab' grew from hanging drops of 5 mg/mL protein with 1.0 M ammonium sulfate at pH 5.8 as precipitant and grew as needles. Complexes were co-crystallized by adding a 3:1 ratio of peptide ELDKWAS to protein and incubating overnight before setting up as hanging drops of 5 mg/mL complex at pH 5.8, using 1.6 M ammonium sulfate at pH 7.0 as precipitant. The crystals grew in two days as large square bipyramids.

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The sequence of the heavy and light variable domains has recently been published (ref. 10) and agrees with the one used in this study with a single correction at amino acid H110, which is a serine rather than a proline as originally stated. The full amino acid sequences of the variable and constant domains of the

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Fab' fragment are shown in Table 1 below (SEQ ID Nos: 6 and 7).

Crystals of the free Fab' belong to the space group P2,2,2, (unit cell: a=63.6 Å; b=76.4 Å; c=94.7 Å) and grow as needles. Crystals of the complex also adopt space group P2,2,2, (unit cell: a=59.0 Å; b=65.0 Å; c=175.6 Å) and grow as square bipyramids. Crystals were flash frozen for data collection. Data were collected on a Rigaku FR-C equipped with Molecular Structure Corp mirror optics and with a Mar345 image plate detector (Fab'2F5) and at the National

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20 Example 2

were processed using DENZO and SCALEPACK (HKL Research).

Synchrotron Light Source in Brookhaven using a Mar30 detector (complex). Data

This Example describes the solution of the structure of the Fab'2F5 complexed and uncomplexed.

The structure of the Fab'2F5 complex was solved by molecular replacement (ref. 24) using PDB entry 1CLZ (ref. 25) minus sidechains and hypervariable loops as the search model. Constant and variable regions were used as independent models. The correct solution had a correlation coefficient of 35.3 (R=47.3%) using data to 3.3 Å. The CNS package (ref. 26) was used for refinement. A 2F_O-F_C map generated after rigid body refinement of the polyalanine model allowed placement of most sidechains. After a cycle of

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simulated annealing, the hypervariable loops were included. Density for the

peptide was clear at this point and could be fitted unambiguously. Following another cycle of annealing, positional and B-factor refinement, waters were

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included where peaks of >3.5 were found in a difference map at an appropriate distance from a donor or acceptor atom.

The structure of the uncomplexed Fab'2F5 was solved by molecular replacement using the refined Fab'2F5 complex minus peptide as the search model. Correlation coefficient was 53.7, R=39.0%. Refinement followed the same procedure as for the complex. Statistics of data collection, processing and structure refinement are given in Table 2 below. The coordinates of the crystal structures have been deposited on April 9, 1999 in the Brookhaven Protein Data Bank under Accession Nos. 2F5A for the uncomplexed structure and 2F5B for the Fab'2F5-epitope complex.

Example

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This Example demonstrates the utility of the three-dimensional structural information of Katinger's epitope (Examples 1 and 2) in the rational design of constraint peptide-based vaccines.

1. ECDKWCS CLP-634 (SEQ ID No: 3)

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Based on the structural information, the Katinger's epitope may be locked with a disulfide bridge between positions 2 and 6 in the peptide ECDKWCS (CLP-634).

The linear peptide ECDKWCS was synthesised manually, on PAM support, by using a standard Solid Phase Peptide Synthesis methodology, with a t-Boc strategy. The crude peptide was cleaved off the resin by high-HF procedure. The crude material (54 mg) was dissolved in methanol (500 mL). 50 mM iodine in methanol was added dropwise, with stirring, until solution became pale-yellow. After 1 min of stirring, Dowex IX2-200 (acetate) resin (approx. 9 g) was added.

The stirring was continued until solution became colourless. The resin was filtered

25 The stirring was continued until solution became colourless. The resin was filtered off, 50 ml of water was added, the mixture was concentrated in vacuo, frozen and lyophilised. The crude cyclic peptide was purified by RP-HPLC.

EdapDKWES CLP-1309 (SEQ ID No: 4)

Based on the structural information, the Katinger's peptide also may be constrained with a lactam bond between positions 2 and 6 in the peptide

30 constrained with a lactam bond between positions 2 and EDapDKWES (CLP-1309).

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The peptide: t-Boc-Glu(OBzl)-Dap(Fmoc)-Asp(OBzl)-Lys(2Cl-Cbz)-Trp(For)-Glu(OFm)-Ser(Bzl)-RESIN was assembled on a PAM solid support. Sidechains of Dap(2) and Glu(6) were subsequently deprotected by treatment with 25% piperidine. The sidechain cyclization was performed on the resin by adding four equivalents of HBTU and 8 equivalents of DIEA and shaking the mixture overnight. The cyclic peptide was cleaved off the resin by a standard HF procedure and the crude product was purified by RP-HPLC.

Abbreviations used in this Example are:

Dap = diaminopropionic acid

 $HBTU = O\text{-}Benzotriazolyl\text{-}N,N,N',N'\text{-}tetramethyluronium}$

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Hexafluorophosphate

DIEA = Di-isopropylethylamine

PAM = 4-Hydroxymethyl-phenylacetamidomethyl resin

I - Delicy

2-Cl-Cbz = 2-Chlorobenzyloxycarbonyl

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For = Formyl

t-Boc = t-Butloxycarbonyl

Fmoc = Fluorenylmethoxycarbonyl

Fm = Fluorenylmethyl

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Both peptides CLP-634 and CLP-1309 were found to be capable of forming an immuno-complex with monoclonal antibody 2F5 and were subsequently co-crystallized with the Fab' fragment. These results indicated that the constrained peptides may mimic the Katinger's epitope and would be useful as peptide-based vaccines.

Example 4

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This Example demonstrates the formation of constrained peptide-carrier conjugates, for use as anti-HIV vaccines.

In order to conjugate the constrained peptide CLP-1309 (Example 3) to a carrier protein, a tetra-peptide Cys-Gly-Gly-Gly was linked to CLP-1309 at the N-

30 terminal end and the resultant peptide was named as CLP-1491. Similarly, a tetrapeptide Gly-Gly-Gly-Cys was linked to CLP-1309 at the C-terminal end, and so the resultant peptide was named as CLP-1492.

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Fifty microlitre of m-maleimidobenzoyl-N-hydroxysuccinimide (MBS, Pierce, 2 mg; 6.3 mmol in 1 mL of tetrahydrofuran or methanol) was added to a protein solution (approximately 10 mg of Hin47 or tetanus toxoid in 2 mL of 0.1 M phosphate buffer, pH 7.5). The reaction mixture was stirred for 30 min at room temperature under argon. The reaction mixture was applied to a Sephadex G-25 column (20 x 300 mm) equilibrated with 20 mM ammonium bicarbonate buffer, pH 7.2 and eluted with the same buffer. Elution was monitored by absorbance at 230 nm, and the eluted protein peak was pooled. The number of maleimide groups incorporated into the carrier was determined by adding excess 2-mercaptoethanol to the activated carrier-MBS and back-titrating the excess using a modified Ellman's method (ref. 31).

A general protocol for peptide-carrier conjugates has been described (ref. 32). Briefly, synthetic peptide (1 mg/mL) in degassed PBS buffer, pH 7.5 mixed with carrier-MBS (1 mg/mL). The reaction mixture was stirred overnight at room temperature under argon atmosphere. Excess N-ethyl-maleimide (Aldrich) was added to quench the reaction, and stirring continued for an additional hour. The insoluble precipitate was filtered off, and the filtrate was subjected to gel filtration chromatography using a Sephadex G-25 column. The peptide-carrier conjugate was collected. The molar ratio of carrier to peptide was determined by using amino acid analysis.

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SUMMARY OF DISCLOSURE

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In summary of this disclosure, the crystal structure of the Fab'2F5 fragment has been elucidated, both in uncomplexed form and complexed with the epitope ELDKWAS, and peptides synthesized to correspond to the constrained structure of the peptide-protein interactions. Modifications are possible within the scope of this invention.

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ALQLTOSPSS LSASVGDRIT ITCRASQGVT SALAWYRQKÞ GSFÞOLLIYD ASSLESGVÞS RESGSGGGTE FTLTISTLRÞ EDFATYYCQQ LHFYÞHTFGG GTRVDVRRTV AAÞSVFIFÞÞ SDEQLKSGTA STVCLLNNFY ÞREAKVÓWKV DNALQSGNSQ ESVTEQDSKD STYSLSSTLT LSKADYEKHK VYACEVTHQG LSSPVTKSFN RGEC (SEQ ID No.: 6)

RITIKESGPP LVKPTQTUIL TCSFSGPSLS DFGVGVGWIR QPPGKALEWL ALIYSIDDEK YSPSLNTRLI ITKDISKNOV VLJWRRVSBV DTRYTECHRR AGITYLGGVF YSPSLNTRLI ITKDISKNOV VLJWRRVSBV DTRYTECHRR RGPTILFGVF LARGRVNAMD VWGQGITVTI SSASTKGPSV FPLAPSSKSI SGGTAALGCL VKDYFBFEPT VSHWISGALTS GVHTFFPAULD SGGLYSLSSV VTVPSSSLGI QTYLICVHRHK PSHTKVDKKV EPKSCDKTHT CPFCPAPELL GGPSVFLFP KFKUTLMSK TFFNTCVVVD VSHEDDEVKE WYVDGVECH NAVTKPBEED NSITIRVUSV LTVLHQDMLN GEFYKCTVMS KAFPAPIEKT ISKAKGOPER PQVTLPPSR DELTKIQVUSL TCLVKGFYPS DIAVEMESNO QPENNYKTTP PVLDSDGSFF LYSKLTVDKS RWQQGNVFSC SVMHEALENH YTOKSLSISP GK (SEQ ID No.: 7)

Table 2
Data Collection, Processing and Structure Refinement Parameters

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<u>Table 3</u> 17

ATOM	HOTA	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	ATOM	HOTA	ATOM	ATOM	H MOTA	H ATOM	H ATOM	ATOM H	ATOM H	ATOM
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4.809	4.579	2.337	3.363	5.674	3.425	4.569	4.626	3.486	3.884	2.824	3.292	2.379	3.831	2.748	2.179	3.446	3.504	1.395	2.229	1.956	1.719	1.731	. 223	. 979	2.361	1.135	049
9 45.388	9 45.001	7 45.128	3 44.906	1 50.910	50.676	50.179	48.941	48.026	46.772	45.673	44.483	43.355	42.026	42.194	41.876	41.808	41.224	41.316	40.897	40.169	41.072	40.229	41.419	40.460	39.794	39.444	39.377
87.678	86.304	86.460	85.805	82.055	81.684	82.133	82.623	82.712	83.478	83.507	84.354	84.306	83.207	83.773	88.301	87.931	86.746	87.319	86.336	85.059	83.841	82.660	81.490	81.598	79.633	80.483	79.646
1.00 24.42	1.00 23.46	1.00 22.03	1.00 22.74	1.00 23.15	1.00 22.75	1.00 22.62	1.00 22.59	1.00 22.45	1.00 22.62	1.00 22.31	1.00 22.26	1.00 21.79	1.00 21.32	1.00 21.64	1.00 20.95	1.00 20.64	1.00 21.12	1.00 20.90	1.00 21.04	1.00 21.35	1.00 21.17	1.00 21.37	1.00 21.06	1.00 21.53	1.00 21.47	1.00 21.70	1.00 21.77

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53.957	53.782	53.434	52.368	51.681	53.2	52.194	50.094	49.948	50.935	50.645	51.6	51,488	52.208	51.366	50.916	52.973	51.756	52.362	50.107	1.596	51.813	51.132	51.354	49.242	50.384	51.116	49.399	50.723
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53.678 54.441

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11.071	9.826	9.327	7.198	8.022	7.472	6.401	6.730	7.565	8.700	8.808	9,239	9.921	10.028	10.432	11.212	11.132	11.588	11.655	12.367	11.632	9.746	10.353	12.029	9.831	10.940	10.971	9.850	10.337
1 49.697	6 48.813	48.824	48.513	48.696	51.255	50.266	50.062	48.764	48.710	46.495	47.534	48.435	47.087	47.497	49.041	48.763	49.036	49.456	50.768	51.620	51.462	51.901	58.835	58.339	58.104	57.131	56.258	55.009
7 82.338	82.455	83.826	83.166	84.066	85.150	87.199	85.719	85.531	86.446	86.252	86.734	89.838	89.119	87.700	89.362	87.918	85.738	86.897	87.203	88.122	86.920	87.892	B5.702	85.217	85.916	86.821	87.132	87.853
1.00 21.90	1.00 21.64	1.00 21.63	1.00 22.38	1.00 22.08	1.00 20.99	1.00 21.48	1.00 21.84	1.00 22.26	1.00 22.92	1.00 23.75	1.00 24.10	1.00 26.45	1.00 25.85	1.00 25.02	1.00 25.99	1.00 25.66	1.00 25.97	1.00 26.06	1.00 26.56	1.00 27.08	1.00 27.45	1.00 27.85	1.00 29.08	1.00 28.91	1.00 29.34	1.00 29.19	1.00 29.05	1.00 28.97

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ATOM ATOM H ATOM H ATOM ATOM ATOM ATOM ATOM ATOM ATOM н АТОМ 2549 CA 2545 ND2 ASN H 117 2552 2550 CB 2548 N 2547 0 2546 C 2544 OD1 2556 CG 2553 2551 C 2543 CG 2555 CB 2557 SD 2554 CA 2559 C 2558 CE 2564 2563 2562 2560 0 2568 0 2567 C 2565 OD1 z 0 z OD2 ASP H 120 ദ B ASN H 117 ASN H 117 ALA H 118 ALA H 118 ASN H 117 ALA H 118 ALA H 118 MET H 119 MET H 119 MET H 119 ALA H 118 MET H 119 MET H 119 MET H 119 ASP H 120 ASP H 120 ASP H 120 MET H 119 ASP H 120 ASP H 120 ASP H 120 ASP H 120 MET H 119 11.673 10.748 10.070 10.093 9.686 8.964 8.984 3.738 6.898 5.413 6.153 7.329 7.209 7.011 5.661 6.806 8.014 7.679 6.894 7.499 6.907 4.880 4.782 8.020 46.691 47.122 51.922 51.173 42.486 45.197 82.333 1.00 21.59 44.511 81.695 1.00 21.58 47.451 81.814 1.00 21.39 44.713 81.466 1.00 21.25 45.364 51.630 40.878 40.826 41.430 41.594 81.542 1.00 21.33 44.767 43.691 79.004 1.00 21.59 42.814 81.012 1.00 21.00 40.807 40.819 40.829 80.773 1.00 21.24 45.836 80.681 1.00 24.35 43.630 40.381 36.027 38.931 81.123 1.00 21.19 83.115 1.00 22.26 81.396 82.858 1.00 21.43 80.053 1.00 22.00 81.716 1.00 21.30 82.116 85.840 83.500 80.759 1.00 21.14 87.075 85.330 1.00 21.92 79.712 83.688 1.00 21.12 84.932 1.00 21.73 83.499 1.00 21.27 1.00 22.65 1.00 22.54 1.00 21.62 1.00 21.35 1.00 22.35 1.00 21.94 1.00 21.67

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Table 4

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79.496 1.00 36.51

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> 5.235 3.676 4.583

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81.668 1.00 26.53

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80.868

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ATOM

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1.00 37.23 1.00 36.76 1.00 1.00 1.00 72.48

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														61.637	61.808	61.727	63.854	62.531	61.700	60.336	60.000	59.597	58.286	58.192	57.305	56.695	56.618	52.924	54.064	52.044	51.627	52.292	54.340	52.319	53.454	53.415	54.431	55.724
														85.617	85.062	84.765	83.541	83.064	83.282	82.795	83.838	83.117	81.026	82.512	83.157	84.834	84.264	88.769	88.891	87.724	85.657	85.041	87.948	86.781	86.870	85.744	85.387	84.829
														1.00 92.11	1.00 53.79	1.00 53.52	1.00 91.74	1.00 91.37	1.00 53.25	1.00 52.63	1.00 49.16	1.00 49.00	1.00 39.87	1.00 48.51	1.00 47.84	1.00 42.32	1.00 42.36	1.00 45.34	1.00 45.30	1.00 45.22	1.00 45.13	1.00 45.27	1.00 45.31	1.00 45.24	1.00 45.24	1.00 45.32	1.00 45.39	1.00 41.97
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CLAIMS

What we claim is:

- An isolated crystal of the Fab' fragment of monoclonal antibody 2F5.
- The isolated crystal of claim 1 consisting of molecules having the threedimensioned structure represented by Figure 1.
- The isolated crystal of claim 1 consisting of molecules having the parameters defined in Table 2.
- 4. The isolated crystal of claim 3 consisting of molecules having a space group P2,2,2,, with said cell dimensions $\underline{a}=63.6$ Å, $\underline{b}=76.4$ Å and $\underline{c}=93.4$ Å.
- 5. The isolated crystal of claim 1 consisting of molecules having a third hypervariable (V3) loop of the heavy chain comprising amino acid residues H98 to H120, as seen in Table 1, having a three-dimensional structure as shown in
- 6. The isolated crystal of claim 5 consisting of molecules wherein said V3 loop has the atomic coordinates shown in Table 3.
- The isolated crystal of claim 1 consisting of molecules having the atomic coordinates deposited with the Protein Data Bank under Accession number 2F5A.
- 8. An isolated crystal of the Fab' fragment of monoclonal antibody 2F5 complexed with a peptide having the amino acid sequence ELDKWAS (SEQ ID No.: 1) or a functional analog thereof.
- The isolated crystal of claim 8 consisting of molecules having a structure at the binding site of the peptide to the Fab' fragment as shown in Figure 3.
- 10. The isolated crystal of claim 8 consisting of molecules having the parameters defined in Table 2.
- 11. The isolated crystal of claim 10 consisting of molecules having a space group P2,2,2,, with unit cell dimensions $\underline{a} = 58.0 \text{ Å}$, $\underline{b} = 65.0 \text{ Å}$ and $\underline{c} = 175.6 \text{ Å}$.
- 12. The isolated crystal of claim 8 wherein said functional analog of said amino acid sequence ELDKWAS is selected from the group consisting of one in which lysine is replaced by arginine and one in which tryptophan is replaced by an amino acid selected from the group consisting of tyrosine, phenylalanine and uncharged histadine.
- The isolated crystal of claim 8 wherein said peptide is ELDKWAS.

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- 14. The isolated crystal of claim 13 wherein the Fab'2F5: ELDKWAS complex has the atomic coordinates of Table 4.
- 15. The isolated crystal of claim 8 wherein said peptide is ELDRWAS (SEQ
- 16. The isolated crystal of claim 15 wherein said Fab2F5: ELDRWAS complex has the atomic coordinates of Table 5.
- 17. The isolated crystal of claim 8 consisting of molecules having the atomic coordinates deposited with the Protein Data Bank under Accession number 2F5B.
- 18. A synthetic peptide which binds to monoclonal antibody 2F5 and which is constrained to provide a three-dimensional structure corresponding to that for the peptide ELDKWAS (SEQ ID No.: 1) shown in Figure 3.
- 19. The synthetic peptide of claim 18 which contains the amino acid sequence DKW or a functional analog thereof constrained in the slightly distorted B-turn configuration of said three-dimensional structure with the tryptophan and lysine sidechains stacked and parallel.
- 20. The synthetic peptide of claim 19 wherein at least one of said tryptophan and lysine amino acids is substituted by an amino acid which retains the peptide-protein interactions shown in Figure 3.
- 21. The synthetic peptide of claim 20 wherein said lysine residues is replaced
- 22. The synthetic peptide of claim 20 wherein said tryptophan is replaced by tyrosine, phenylalanine or uncharged histadine.
- 23. The synthetic peptide of claim 20 wherein said lysine residue is replaced by arginine and said tryptophan is replaced by tyrosine, phenylalanine or uncharged histadine.
- 24. The synthetic peptide of claim 19 wherein said peptide contains a disulphide bridge to maintain said amino acid sequence DKW or functional analog thereof in the respective orientation of the amino acid residues as shown in Figure
- 25. The synthetic peptide of claim 24 wherein said peptide has the amino acid sequence ECDKWCS (SEQ ID No.: 3) and said disulphide bridge is established between said cysteine (C) residues.

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- 26. The synthetic peptide of claim 19 wherein said peptide contains a lactain bond to maintain said amino acid sequence DKW or functional analog thereof in the respective orientation of the amino acid residues as shown in Figure 3.
- 27. The synthetic peptide of claim 26 wherein said peptide has the formula EDapDKWES (SEQ ID No.: 5) and said lactam bond is formed between the Dap and glutamate (E) residues.
- The synthetic peptide of claim 18 which is linked to a carrier protein
- A method of making a peptide binding to monoclonal antibody 2F5, which imprises:

co-crystallizing a Fab' fragment of the monoclonal antibody 2F5 with a peptide having the amino acid sequence ELDKWAS (SEQ ID No: 1) or functional analog thereof to form a crystalline complex,

analyzing the crystalline complex to determine the three-dimensional orientation of the bound peptide in relation to the Fab' fragment, and

synthesizing a peptide containing at least amino acids DKW or functional analogs thereof constrained in the determined three-dimensional orientation.

30. The method of claim 29 wherein said functional analog of the peptide having the amino acid sequence ELDKWAS has the amino acid sequence ELDRWAS (SEQ ID No.: 2).

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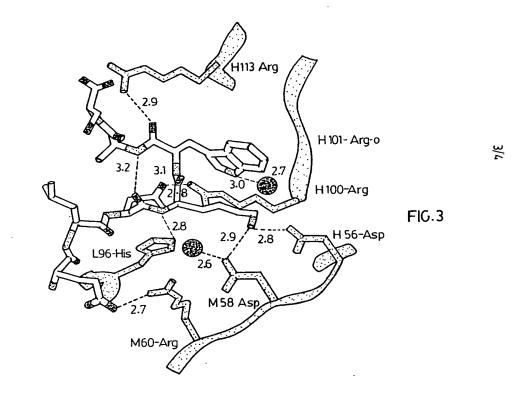
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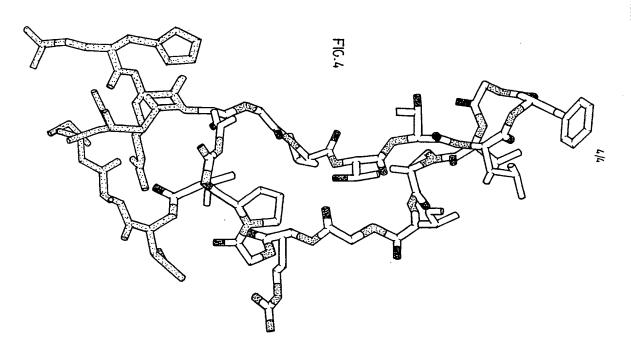
FIG. 2

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FIG.1





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